

Oral Presentation

Open Access

Enhanced protein expression through strain selection, gene disruption, improved vector design and co-expression of endogenous chaperones

Darrell Sleep*, Chris Finni and Leslie Evans

Address: Novozymes Delta Ltd, Castle Court, 59 Castle Boulevard, Nottingham, UK

* Corresponding author

from The 4th Recombinant Protein Production Meeting: a comparative view on host physiology
Barcelona, Spain. 21–23 September 2006

Published: 10 October 2006

Microbial Cell Factories 2006, **5**(Suppl 1):S29 doi:10.1186/1475-2859-5-S1-S29

© 2006 Sleep et al; licensee BioMed Central Ltd.

Background

The use of *Saccharomyces cerevisiae* as a host system has been limited by the perception of limited secretion capacity, unstable episomal vectors and aberrant glycosylation. Solutions to all of these limitations are now available.

Results

An analysis of a series of haploid laboratory yeast strains revealed significant intra-strain variability and unstable plasmid segregation. By combining classic chemical mutagenesis and selection a family of highly efficient *Saccharomyces cerevisiae* strains has been developed for the commercial production of biopharmaceutical products. When combined with a stable [1], high copy number [2], episomal expression vector system and a strong constitutive promoter, secreted recombinant protein expression titres in excess of 4 g/L were achieved (see Figure 1). Specific genetic modifications to the host were also introduced to increase product yield and control post-translational modifications, such as proteolysis and glycosylation.

The expression vectors have been further enhanced to facilitate the stable co-expression of multiple proteins. When one of these proteins is a chaperone, the titre of co-expressed recombinant transferrin was increased 15-fold. The applicability of this system has been demonstrated with a wide range of heterologous proteins and is scalable from 10 mL shake flask to cGMP manufacture at high cell density fermentation (8,000 L) in a defined synthetic

medium; designed to be integrated with cost-efficient downstream processing.

Conclusion

Significant intra-strain variability and unstable episomal plasmid systems have limited the usefulness of *Saccharomyces cerevisiae* as an industrial host for the production of biopharmaceuticals. However co-enhancement of the episomal vector system and the host strains is not only possible but has led to significant improvements in recombinant protein production.

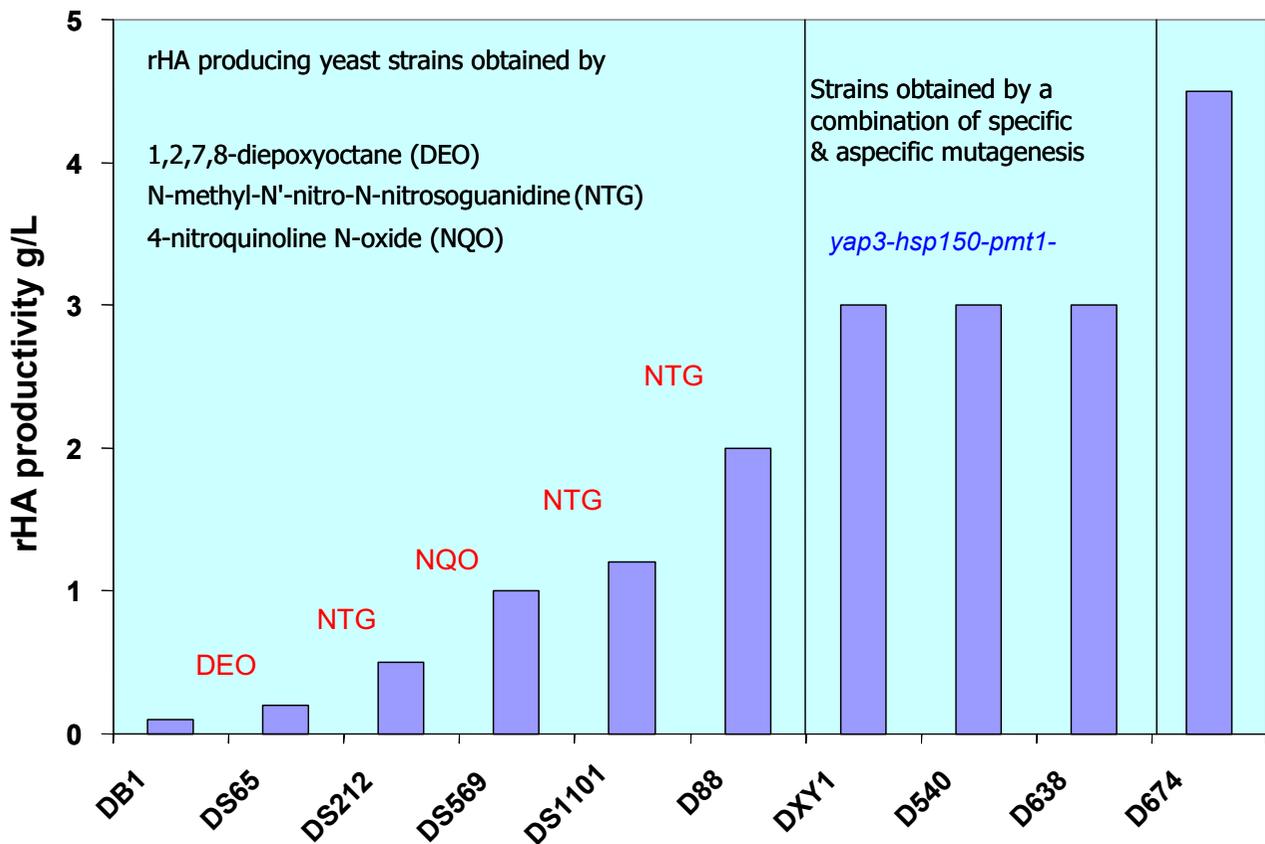


Figure 1
Enhancement of protein production through chemical mutagenesis and specific gene disruptions.

References

1. Chinery SA, Hincliffe E: **A novel class of vector for yeast transformation.** *Curr Genet* 1989, **16**:21-25.
2. Sleep D, Finnis C, Turner AJ, Evans LR: **Yeast 2 mm plasmid copy number is elevated by a mutation in the nuclear gene *UBC4*.** *Yeast* 2001, **18**:403-421.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

